

This article was downloaded by:

On: 22 January 2011

Access details: *Access Details: Free Access*

Publisher *Taylor & Francis*

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



## Journal of Asian Natural Products Research

Publication details, including instructions for authors and subscription information:

<http://www.informaworld.com/smpp/title~content=t713454007>

### New long-chain hydroxyalkyl ferulates from the root bark of *Lycium chinense* Mill.

Shu Pan<sup>a</sup>; Ai-Jun Hou<sup>a</sup>

<sup>a</sup> Department of Pharmacognosy, School of Pharmacy, Fudan University, Shanghai, China

**To cite this Article** Pan, Shu and Hou, Ai-Jun(2009) 'New long-chain hydroxyalkyl ferulates from the root bark of *Lycium chinense* Mill.', *Journal of Asian Natural Products Research*, 11: 7, 681 – 685

**To link to this Article:** DOI: 10.1080/10286020903002139

**URL:** <http://dx.doi.org/10.1080/10286020903002139>

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: <http://www.informaworld.com/terms-and-conditions-of-access.pdf>

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

## New long-chain hydroxyalkyl ferulates from the root bark of *Lycium chinense* Mill.

Shu Pan and Ai-Jun Hou\*

Department of Pharmacognosy, School of Pharmacy, Fudan University, Shanghai 201203, China

(Received 5 March 2009; final version received 27 April 2009)

Two new long-chain hydroxyalkyl ferulates, (*E*)-22-hydroxydocosyl 3-(4-hydroxy-3-methoxyphenyl)acrylate (lyciumol A, **1**) and (*E*)-20-hydroxyeicosyl 3-(4-hydroxy-3-methoxyphenyl)acrylate (lyciumol B, **2**), together with eight known compounds, were isolated from the root bark of *Lycium chinense* Mill. Their structures were elucidated by spectroscopic methods, including 1D, 2D NMR, and HR-ESI-MS analysis.

**Keywords:** *Lycium chinense*; long-chain hydroxyalkyl ferulates; lyciumol A; lyciumol B

### 1. Introduction

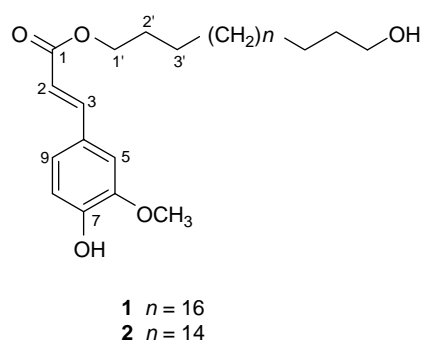
*Lycium chinense* Mill. is a deciduous shrub widely distributed over China. Its dried root bark is used as traditional Chinese medicine ‘Cortex Lycii’ for the treatment of night sweat, cough, non-traumatic hemorrhage, bloody stranguria, hypertension, and ulcer. The modern pharmacological study showed that the crude extract from the root bark of *L. chinense* had hypotensive, hypoglycemic, antipyretic, and anti-stress ulcer activities in experimental animals [1]. A limited number of glycosides [2], cyclic peptides [3], terpenes [4], spermine alkaloid [5], cerebroside [6], and phenolic amides [7,8] were isolated from this plant, some of these compounds exerting hypotensive, anti-hepatotoxic, anti-oxidative, and anti-fungal effects. Further phytochemical investigations on the root bark of *L. chinense* by our group afforded two new long-chain hydroxyalkyl ferulates, lyciumols A–B (**1–2**) (Figure 1), and eight known compounds, aurantiamide acetate (**3**), scopoletin (**4**), scoparone (**5**), vanillic acid (**6**), vanillin (**7**),

isovanilin (**8**), *p*-hydroxybenzoic acid (**9**), and daucosterol (**10**). To our knowledge, compounds **1** and **2** are the first two ferulates bearing a long-chain hydroxyalkyl group from the plant genus *Lycium*, although ferulates being found quite commonly. Moreover, compounds **5** and **7–9** were isolated from the genus *Lycium* for the first time. In this paper, we mainly describe the structural elucidation of compounds **1** and **2**.

### 2. Results and discussion

Lyciumol A (**1**), a white amorphous powder, was assigned a molecular formula of C<sub>32</sub>H<sub>54</sub>O<sub>5</sub> by HR-ESI-MS at *m/z* 541.3884 [M+Na]<sup>+</sup>. The IR spectrum of **1** exhibited absorptions for hydroxyl groups (3431 cm<sup>-1</sup>), a carbonyl group (1711 cm<sup>-1</sup>), and an aromatic ring (1602 and 1469 cm<sup>-1</sup>). The UV absorption maxima at 321 and 241 nm resembled those of ferulic acid derivatives [9]. The <sup>1</sup>H NMR spectrum (Table 1) showed signals of an aromatic *ABX* spin system

\*Corresponding author. Email: ajhou@shmu.edu.cn

Figure 1. Structures of compounds **1** and **2**.

at  $\delta_{\text{H}}$  7.07 (1H, dd,  $J = 1.5, 8.1$  Hz), 7.04 (1H, br d,  $J = 1.5$  Hz), and 6.92 (1H, d,  $J = 8.1$  Hz), a *trans*-double bond conjugated with a carbonyl group at  $\delta_{\text{H}}$  7.61 and 6.29 (each 1H, d,  $J = 16.0$  Hz), and a methoxy group at  $\delta_{\text{H}}$  3.92 (3H, s). These data suggested the presence of a *trans*-ferulic acid group by comparison of the data with those reported previously [10]. Furthermore, the  $^1\text{H}$  NMR spectrum also provided signals of two downfield methylene groups at  $\delta_{\text{H}}$  4.18 and 3.64 (each 2H,

$J = 6.6$  Hz), two upfield methylene groups at  $\delta_{\text{H}}$  1.68 and 1.55 (each 2H, m), and 36 overlapped protons at  $\delta_{\text{H}}$  1.24–1.41, corresponding to an oxygenated long-chain alkyl group. The  $^{13}\text{C}$  NMR spectrum (Table 1) exhibited a carbonyl signal at  $\delta_{\text{C}}$  167.4, eight  $\text{sp}^2$ -hybridized carbon signals at  $\delta_{\text{C}}$  109.3–147.4, and a methoxy group signal at  $\delta_{\text{C}}$  55.9, which were associated with the ferulic acid moiety [10]. In addition, two downfield methylene signals at  $\delta_{\text{C}}$  64.6 and 63.0 and carbon signals at  $\delta_{\text{C}}$  25.7–32.8 were attributed to the oxygenated long-chain alkyl group, which was deduced to be a hydroxydodecyl group by the molecular formula of **1**. Analysis of HMQC and HMBC spectra assigned all  $^1\text{H}$  and  $^{13}\text{C}$  NMR signals (Table 1). The methoxy group at  $\delta_{\text{H}}$  3.92 was located at C-6, as established by its HMBC correlation with C-6 ( $\delta_{\text{C}}$  146.8). The methylene groups at  $\delta_{\text{H}}$  4.18 and 3.64 were assigned to be  $\text{CH}_2$ -1' and  $\text{CH}_2$ -22', respectively, according to their downfield chemical shifts and HMBC correlations

Table 1.  $^1\text{H}$  (500 MHz) and  $^{13}\text{C}$  (125 MHz) NMR spectral data of **1** and **2** (in  $\text{CDCl}_3$ ,  $\delta$  in ppm,  $J$  in Hz).

Position	<b>1</b>		<b>2</b>	
	$\delta_{\text{H}}$	$\delta_{\text{C}}$	$\delta_{\text{H}}$	$\delta_{\text{C}}$
1	–	167.4	–	167.4
2	6.29 (d, $J = 16.0$ )	115.6	6.29 (d, $J = 16.0$ )	115.7
3	7.61 (d, $J = 16.0$ )	144.7	7.61 (d, $J = 16.0$ )	144.6
4	–	127.0	–	127.1
5	7.04 (br d, $J = 1.5$ )	109.3	7.03 (br d, $J = 1.2$ )	109.3
6	–	146.8	–	146.9
7	–	147.4	–	147.9
8	6.92 (d, $J = 8.1$ )	114.7	6.92 (d, $J = 8.2$ )	114.7
9	7.07 (dd, $J = 1.5, 8.1$ )	123.0	7.08 (dd, $J = 1.2, 8.2$ )	123.1
6-OMe	3.92 (s)	55.9	3.93 (s)	56.0
1'	4.18 (t, $J = 6.6$ )	64.6	4.19 (t, $J = 6.6$ )	64.6
2'	1.68 (m)	28.7	1.68 (m)	28.8
3'	1.24–1.41 (overlap)	26.0	1.24–1.40 (overlap)	26.0
4'–17'	1.24–1.41 (overlap)	29.3–29.7	1.24–1.40 (overlap)	29.3–29.7
18'	1.24–1.41 (overlap)	29.3–29.7	1.24–1.40 (overlap)	25.8
19'	1.24–1.41 (overlap)	29.3–29.7	1.55 (m)	32.8
20'	1.24–1.41 (overlap)	25.7	3.64 (t, $J = 6.6$ )	63.1
21'	1.55 (m)	32.8	–	–
22'	3.64 (t, $J = 6.6$ )	63.0	–	–

from CH<sub>2</sub>-1' ( $\delta_{\text{H}}$  4.18) to C-1 ( $\delta_{\text{C}}$  167.4). The methylene groups at  $\delta_{\text{H}}$  1.68 and 1.55 were determined to be CH<sub>2</sub>-2' and CH<sub>2</sub>-21', respectively, as supported by HMBC correlations from CH<sub>2</sub>-2' ( $\delta_{\text{H}}$  1.68) to C-1' ( $\delta_{\text{C}}$  64.6) and C-3' ( $\delta_{\text{C}}$  26.0) and from CH<sub>2</sub>-21' ( $\delta_{\text{H}}$  1.55) to C-22' ( $\delta_{\text{C}}$  63.0) and C-20' ( $\delta_{\text{C}}$  25.7) (Figure 2). Thus, the structure of **1** was elucidated as (*E*)-22-hydroxydocosyl 3-(4-hydroxy-3-methoxyphenyl)acrylate, and named lyciumol A (Figure 1).

Lyciumol B (**2**), a white amorphous powder, was assigned a molecular formula of C<sub>30</sub>H<sub>50</sub>O<sub>5</sub> by HR-ESI-MS at  $m/z$  513.3544 [M+Na]<sup>+</sup>. The UV and IR data of **2** were very similar to those of **1**. Comparison of the <sup>1</sup>H and <sup>13</sup>C NMR spectral data of these two compounds (Table 1) showed that they shared the same ferulyl moiety, but had the different long-chain alkyl groups. Compound **2** contained a hydroxyeicosyl group rather than the hydroxydocosyl group in **1**, as supported by the molecular formula of **2**. Thus, the structure of **2** was elucidated as (*E*)-20-hydroxyeicosyl 3-(4-hydroxy-3-methoxyphenyl)acrylate, and named lyciumol B (Figure 1).

Eight known compounds were isolated and identified as aurantiamide acetate (**3**) [11], scopoletin (**4**) [12], scoparone (**5**) [13], vanillic acid (**6**) [14], vanillin (**7**) [15], isovanillin (**8**) [15], *p*-hydroxybenzoic

acid (**9**) [16], and daucosterol (**10**) [16] by comparison of the spectroscopic data with those reported previously.

### 3. Experimental

#### 3.1 General experimental procedures

The UV spectra were determined on a Shimadzu UV-2401PC spectrophotometer and IR spectra were measured on a Nicolet Avatar-360 spectrometer. The 1D and 2D NMR spectra were recorded on Bruker DRX-500 and Varian Mercury Plus 400 instruments, using residual solvent peaks of CDCl<sub>3</sub> ( $\delta_{\text{H}}$  7.27,  $\delta_{\text{C}}$  76.9) as the standard. Chemical shifts ( $\delta$ ) were reported in ppm and coupling constants (*J*) in Hz. EI-MS measurements were carried out on a VG Auto Spec-3000 spectrometer and HR-ESI-MS data were obtained on an AB QSTAR Pulsar mass spectrometer. Column chromatography was performed on silica gel H (10–40  $\mu\text{m}$  and 200–300 mesh; Institute of Chemical Technology, Yantai, China), Diaion HP-20 (Mitsubishi Chemical Co., Tokyo, Japan), MCI gel CHP-20P (75–150  $\mu\text{m}$ ; Mitsubishi Chemical Co.), and Chromatorex RP-18 gel (20–45  $\mu\text{m}$ ; Fuji Silysia Chemical, Ltd, Kasugai, Japan). TLC analysis was run on precoated silica gel GF<sub>254</sub> plates (10–40  $\mu\text{m}$ ; Institute of Chemical Technology). Preparative HPLC was performed on Agilent 1200 (Agilent Technologies, Santa Clara, CA, USA) and Sepax Amethyst C<sub>18</sub> column (10.0  $\times$  150 mm, 5  $\mu\text{m}$ ; Sepax Technologies, Inc., Newark, NJ, USA).

#### 3.2 Plant material

The dried root bark of *L. chinense* Mill. was collected from Hebei Province, China, in 2007, and was identified by Dr Yun Kang, Fudan University. A voucher specimen (TCM 2007-07-09 Hou) is deposited in the Herbarium of the Department of Pharmacognosy, School of Pharmacy, Fudan University.

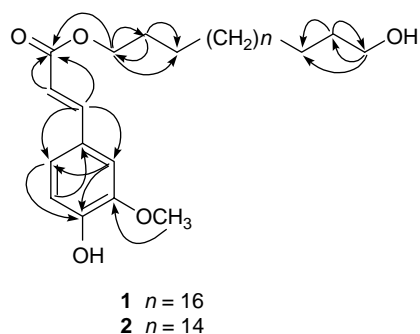


Figure 2. Selected HMBC (H  $\rightarrow$  C) correlations of compounds **1** and **2**.

### 3.3 Extraction and isolation

The dried and powdered root bark of *L. chinense* Mill. (9.5 kg) was percolated with 80% ethanol at room temperature. The filtrate was evaporated *in vacuo* to give a residue, which was suspended in H<sub>2</sub>O and partitioned successively with CHCl<sub>3</sub> and EtOAc. The CHCl<sub>3</sub> extract (90 g) was subjected to column chromatography (CC) on silica gel eluted with a gradient of petroleum ether–Me<sub>2</sub>CO (50:1 → 1:2) to afford fractions 1–14. Fraction 8 (6 g) was separated by CC on silica gel (petroleum ether–Me<sub>2</sub>CO, 6:1) to give fractions 8.1–8.8. Fraction 8.6 (972 mg) was purified by CC on silica gel (CHCl<sub>3</sub>–Me<sub>2</sub>CO, 100:1) to afford fractions 8.6.1–8.6.9. Fraction 8.6.1 (12 mg) was separated by CC over silica gel (petroleum ether–Me<sub>2</sub>CO, 8:1) to give **6** (3 mg). Fraction 8.6.4 (15 mg) was purified by CC on RP-18 (MeOH–H<sub>2</sub>O, 40:60) to yield **7** (3 mg) and **8** (4 mg). Fraction 8.6.7 (25 mg) was purified by HPLC (flow rate 1 ml/min, UV detector 210 nm) using 98% MeOH to yield **1** (5 mg, *t<sub>R</sub>* 63 min). Fraction 8.6.8 (32 mg) was purified by HPLC (flow rate 1 ml/min, UV detector 210 nm) using 96% CH<sub>3</sub>CN to give **2** (8 mg, *t<sub>R</sub>* 61 min). Fraction 8.7 (82 mg) was separated by CC over silica gel (petroleum ether–Me<sub>2</sub>CO, 5:1) to afford **3** (5 mg). Fraction 10 (602 mg) was chromatographed over silica gel (petroleum ether–Me<sub>2</sub>CO, 4:1), followed by CC on RP-18 (MeOH–H<sub>2</sub>O, 60:40) to give **4** (9 mg). Fraction 12 (112 mg) was separated by CC on MCI gel CHP-20P (MeOH–H<sub>2</sub>O, 60:40 → 90:10) to afford compound **9** (3 mg). Fraction 14 (9 g) was isolated by CC on Diaion HP-20 eluted with a gradient of MeOH–H<sub>2</sub>O (30:70 → 100:0) to give **5** (4 mg) and **10** (1244 mg).

#### 3.3.1 *Lyciumol A (1)*

A white amorphous powder; UV (CHCl<sub>3</sub>)  $\lambda_{\max}$  (log  $\epsilon$ ): 321 (4.05), 241 (3.88) nm; IR

(KBr)  $\nu_{\max}$ : 3431, 2918, 2850, 1711, 1631, 1602, 1517, 1469, 1431, 1377, 847, 816 cm<sup>-1</sup>; <sup>1</sup>H and <sup>13</sup>C NMR spectral data, see Table 1; EI-MS *m/z* (%): 518 (55, [M]<sup>+</sup>), 194 (67), 177 (100), 150 (44), 137 (39), 69 (13), 55 (8); HR-ESI-MS *m/z*: 541.3884 [M + Na]<sup>+</sup> (calcd for C<sub>32</sub>H<sub>54</sub>O<sub>5</sub>Na, 541.3868).

#### 3.3.2 *Lyciumol B (2)*

A white amorphous powder; UV (CHCl<sub>3</sub>)  $\lambda_{\max}$  (log  $\epsilon$ ): 327 (4.07), 236 (3.92) nm; IR (KBr)  $\nu_{\max}$ : 3441, 2918, 2850, 1710, 1631, 1602, 1518, 1469, 1430, 1377, 846, 816 cm<sup>-1</sup>. <sup>1</sup>H and <sup>13</sup>C NMR spectral data, see Table 1; EI-MS *m/z* (%): 490 (55, [M]<sup>+</sup>), 194 (82), 177 (100), 150 (44), 137 (38), 117 (8), 69 (14), 55 (10); HR-ESI-MS *m/z*: 513.3544 [M + Na]<sup>+</sup> (calcd for C<sub>30</sub>H<sub>50</sub>O<sub>5</sub>Na, 513.3555).

### Acknowledgements

The authors thank Prof. Qin-Shi Zhao, Kunming Institute of Botany, Chinese Academy of Sciences, for his kind help. We are also thankful to the Shanghai Institute of Organic Chemistry, Chinese Academy of Sciences for recording the NMR spectra.

### References

- [1] T. Morota, H. Sasaki, M. Chin, T. Sato, N. Katayama, K. Fukuyama, and H. Mitsuhashi, *Shoyakugaku Zasshi* **41**, 169 (1987).
- [2] M. Terauchi, H. Kanamori, M. Nobuso, S. Yahara, and K. Yamasaki, *Nat. Med.* **52**, 167 (1998).
- [3] S. Yahara, C. Shigeyama, T. Ura, K. Wakamatsu, T. Yasuhara, and T. Nohara, *Chem. Pharm. Bull.* **41**, 703 (1993).
- [4] A. Sannai, T. Fujimori, and K. Kato, *Phytochemistry* **21**, 2986 (1982).
- [5] S. Funayama, K. Yoshida, H. Konno, and H. Hikino, *Tetrahedron Lett.* **21**, 1355 (1980).
- [6] S.Y. Kim, Y.H. Choi, H. Huh, J. Kim, Y.C. Kim, and H.S. Lee, *J. Nat. Prod.* **60**, 274 (1997).

- [7] S.H. Han, H.H. Lee, I.S. Lee, Y.H. Moon, and E.R. Woo, *Arch. Pharm. Res.* **25**, 433 (2002).
- [8] D.G. Lee, Y. Park, M.R. Kim, H.J. Jung, Y.B. Seu, K.S. Hahm, and E.R. Woo, *Biotechnol. Lett.* **26**, 1125 (2004).
- [9] H.T. Lu, H.F. Sun, B.H. Qu, and H.Y. Dai, *Chin. J. Anal. Chem.* **35**, 1425 (2007).
- [10] B. D'Abrosca, A. Fiorentino, S. Pacifico, G. Cefarelli, P. Uzzo, M. Letizia, and P. Monaco, *Bioorg. Med. Chem. Lett.* **17**, 4135 (2007).
- [11] J. Tang, S. Tewtrakul, Z.T. Wang, and Z.B. Tu, *J. Chin. Pharm. Sci.* **12**, 231 (2003).
- [12] X.W. Zhou, G.J. Xu, and Q. Wang, *Chin. J. Chin. Mater. Med.* **21**, 675 (1996).
- [13] Y.X. Pan, C.X. Zhou, S.L. Zhang, X.X. Zheng, and Y. Zhao, *J. Chin. Pharm. Sci.* **13**, 92 (2004).
- [14] X.L. Wei and J.Y. Liang, *Chin. Tradit. Herb. Drugs* **34**, 580 (2003).
- [15] L. Wang, Z.Q. Yin, L.H. Zhang, W.C. Ye, X.Q. Zhang, W.B. Shen, and S.X. Zhao, *Chin. J. Chin. Mater. Med.* **32**, 1300 (2007).
- [16] Y.K. Qiu, D.Q. Dou, Y.P. Pei, M. Yoshikawa, H. Matsuda, and Y.J. Chen, *Chin. J. Chin. Mater. Med.* **30**, 1824 (2005).